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PAPER

# Phylogeny and the prediction of tree functional diversity across novel continental settings

Nathan G. Swenson<sup>1\*</sup>, Michael D. Weiser<sup>2</sup>, Lingfeng Mao<sup>1</sup>, Miguel B. Araújo<sup>3,4,5</sup>, José Alexandre F. Diniz-Filho<sup>6</sup>, Johannes Kollmann<sup>7</sup>, David Nogués-Bravo<sup>4</sup>, Signe Normand<sup>8,12</sup>, Miguel Ángel Rodríguez<sup>9</sup>, Raúl García-Valdés<sup>10</sup>, Fernando Valladares<sup>3,11</sup>, Miguel A. Zavala<sup>9</sup> and Jens-Christian Svenning<sup>12</sup>

<sup>1</sup>Department of Biology, University of Maryland, College Park, MD, USA,

<sup>2</sup>Department of Biology, University of Oklahoma, Norman, OK 73019, USA, <sup>3</sup>Museo Nacional de Ciencias Naturales, CSIC, Calle Jose Gutierrez Abascal, 2, Madrid, 28006, Spain, <sup>4</sup>Center for Macroecology, Evolution and Climate, National Museum of Natural Sciences, University of Copenhagen, Universitetsparken 15, Copenhagen, 2100, Denmark,

<sup>5</sup>CIBIO/InBio-UE: Centro de Investigação em Biodiversidade e Recursos Genéticos, Universidade de Evora, Évora, 7000-890, Portugal, <sup>6</sup>Departamento de Ecologia, Universidade Federal de Goiás, Campus II, Goiânia, GO, Brazil, <sup>7</sup>Restoration Ecology, Department of Ecology and Ecosystem Management, Technische Universität München, Emil-Ramann-Str. 6, Freising, 85354, Germany,

<sup>8</sup>Dynamic Macroecology, Swiss Federal Research Institute WSL, Zürcherstr. 111, Birmensdorf, 8903, Switzerland, <sup>9</sup>Forest Ecology and Restoration Group, Department of Life Sciences, Science Building, University of Alcalá, Campus Universitario, Alcalá de Henares (Madrid), 28805, Spain, <sup>10</sup>Centre of Ecological Research and Forestry Applications (CREAF),

Department of Animal Biology, Plant Biology and Ecology, Autonomous University of Barcelona, Barcelona, Spain, <sup>11</sup>Departamento de Biología y Geología, Escuela Superior de Ciencias Experimentales y Tecnológicas, Universidad Rey Juan Carlos, c/Tulipán s/n, Móstoles, E-28933, Spain, <sup>12</sup>Section for Ecoinformatics & Biodiversity, Department of Bioscience, Aarhus University, Ny Munkegade 114, 8000, Aarhus C, Denmark

\*Correspondence: Nathan G. Swenson, Department of Biology, University of Maryland, College Park, MD, USA. E-mail: swenson@umd.edu

## ABSTRACT

**Aim** Mapping the distribution and diversity of plant function is critical for projecting future changes to vegetation under global change. Maps of plant function, however, are scarce due very sparse global trait data matrices. A potential solution to this data limitation is to utilize the known levels of phylogenetic signal in trait data to predict missing values. Here we aim to test existing phylogenetic comparative methods for imputing missing trait data for the purpose of producing continental-scale maps of plant function.

**Location** North America and Europe.

**Methods** Phylogenetic imputation models and trait data from one continent were used to predict the trait values for tree species on the other continent and to produce trait maps. Predicted maps of trait means, variances and functional diversity were compared with known maps to quantify the degree to which predicted trait values could estimate spatial patterns of trait distributions and diversity.

**Results** We show that the phylogenetic signal in plant functional trait data can be used to provide robust predictions of the geographical distribution of tree functional diversity. However, predictions for traits with little phylogenetic signal, such as maximum height, are error prone. Lastly, trait imputation methods based on phylogenetic generalized least squares tended to outperform those based on phylogenetic eigenvectors.

**Main conclusions** It is possible to predict patterns of functional diversity across continental settings with novel species assemblages for most of the traits studied for which we have no direct trait information, thereby offering an effective method for overcoming a key data limitation in global change biology, macroecology and ecosystem modelling.

## Keywords

Forest ecology, imputation, plant biodiversity, phylogeny, temperate forest, trait biogeography.

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## INTRODUCTION

Theoretical and empirical ecological investigations suggest that strong linkages exist between plant functional diversity and ecosystem function (Tilman *et al.*, 1997; Loreau *et al.*, 2001). The distribution of functional diversity across a variety of spatial scales is therefore of fundamental interest to ecosystem modellers. Quantifying the continental-scale distribution of plant functional diversity has, however, been particularly challenging due to limitations in the available species trait data (Reich, 2005; Swenson & Weiser, 2010; Swenson *et al.*, 2012). This lack of information has led ecosystem modellers to characterize vegetation types using a few plant functional types, leading to coarse and potentially inaccurate projections of ecosystem function under global climate change (Purves & Pacala, 2008; van Bodegom *et al.*, 2012).

The most obvious obstacle to estimating the continental-scale distribution of plant functional diversity is the requirement for species-level functional trait data that are linked to performance for thousands of species distributed across vast areas, as well as specific knowledge about how such traits are directly or indirectly linked to ecosystem function or persistence. It may require many years to collect such data, even in less diverse temperate floras, and much longer in highly diverse tropical floras (Swenson, 2013; Umaña *et al.*, 2015). A potentially powerful and more easily employed alternative or stopgap measure is to take advantage of phylogenetic signal in functional traits (i.e. the tendency for closely related species to have similar trait values) to estimate the function of individual species. Plant ecologists have demonstrated a large degree of phylogenetic signal in global-scale studies of plant functional traits (e.g. Moles *et al.*, 2005; Swenson & Enquist, 2007), suggesting that reasonable estimates of trait values for species that are absent in global databases may be possible based on their phylogenetic position. Specifically, phylogenetic imputation, in which a model of trait evolution is applied to a phylogeny to estimate the missing trait values for species, holds tremendous promise (Swenson, 2014a). However, these methods have not yet been applied to large plant trait databases nor have they been used to predict the spatial distribution of multiple traits across continents or to predict the distribution of functional diversity itself.

Here, we show that phylogenetic information can be used to generate robust predictions of the distribution of individual functional traits and the overall functional diversity of tree assemblages on continental scales. The analyses focus on using phylogenetic generalized least squares (pGLS) regression and phylogenetic eigenvector regression to evaluate phylogenetic signal in available trait data from one continent and to estimate the functional trait values of individual species on another continent based upon their phylogenetic position (Martins & Hansen, 1997; Garland & Ives, 2000; Swenson, 2014a,b). The analyses were conducted using the geographical distribution of tree species in eastern North America and Europe, a phylogenetic tree of these species and data for four key functional traits (leaf size, maximum

height, seed mass and wood density) for all species. The specific questions we ask are: (1) can the mean and variance of individual traits and multivariate functional diversity of tree species on one continent be predicted by simply knowing the traits and phylogenetic positions of a different set of species on a different continent; (2) does a lack of detailed phylogenetic information within genera greatly hinder predictive models; and (3) do alternative phylogenetic regression models, such as those built using phylogenetic eigenvectors, provide robust predictions of the distribution and diversity of plant function across continents?

## MATERIALS AND METHODS

### Geographical data

Geographical range maps for 273 eastern North American and 121 European tree species were used in this study (we defined a 'tree' as any free-standing woody plant with a maximum height greater than 10 m). Tree species in these two regions that did not have trait data available in the literature were not included in the study. The eastern North American tree range maps were downloaded from the United States Geological Survey (<http://esp.cr.usgs.gov/data/little/>) and gridded into 1° squares. The European tree range data were digitized from the Atlas Flora Europaeae (<http://www.luomus.fi/english/botany/afe/>) and were gridded using the atlas's map grid system where grid cells are 50 km<sup>2</sup> on average. The two tree floras used are well known for their compositional similarity, making them a probable 'best case scenario' for phylogenetic imputation. Specifically, 72.7% of the genera in our European data set are in the North American data set and 25.2% of the North American genera are in our European data set.

### Phylogenetic tree

A single phylogenetic tree was generated for this study using the eco-informatics software Phylocom (Webb & Donoghue, 2005). Specifically, we used the Phylocom R20100701.new backbone phylogeny and our species list to produce a phylogeny. Generally, the degree of relatedness between species within genera was left unresolved using this approach (i.e. all congeneric species pairs were treated as equally related). We used this approach to generate the phylogenetic tree because it is likely to be the approach most widely employed by ecologists in the future attempting to predict trait data on continental scales, particularly in geographical regions where DNA sequences for most species are unavailable (e.g. tropical floras).

### Trait data

This study utilized data for four traits that indicate where a species falls along the spectrum of plant ecological strategies (e.g. Grubb, 1977; Dolph & Dilcher, 1980; Chave *et al.*, 2009; Moles *et al.*, 2009). These traits were also used because they are widely available, allowing for model testing. The traits we

135 considered were maximum height, seed mass, wood density  
 136 and leaf size, and were recorded for every species (i.e. there  
 137 were no missing trait values for any species or trait). The  
 138 maximum height data came from the literature where we  
 139 recorded the absolute largest value reported (Britton & Sha-  
 140 fer, 1923; Polunin, 1976) and the United States Department  
 141 of Agriculture PLANTS database (<http://plants.usda.gov>). The  
 142 wood density data came from the global wood density data-  
 143 base published by Chave *et al.* (2009) and from additional  
 144 literature sources (Iatsenko-Khmelevski, 1954; Bosshard,  
 145 1974). Leaf area was estimated as the product of the reported  
 146 leaf length, leaf width and 0.70 to account for leaf tapering.  
 147 This calculation has recently been shown to produce values  
 148 that are highly correlated with the known area of leaves  
 149 (Kraft *et al.*, 2008) and represents a pragmatic approach for  
 150 estimating leaf area for hundreds of species from the litera-  
 151 ture. For some species, the leaf length and/or width was not  
 152 available in the literature and was recorded by N.G.S. using  
 153 herbarium specimens in the Gray Herbarium at Harvard  
 154 University and the Michigan State University Herbarium.  
 155 Because the degree of leaf shrinkage across these taxa was not  
 156 known and leaves could not be rehydrated we retained the  
 157 dry dimensions. We expect that this introduced error is mini-  
 158 mal given the total variation in leaf size in our data set and  
 159 would probably bias towards weaker predictions. Seed mass  
 160 was recorded from the Kew Millennium Seed Database  
 161 (<http://data.kew.org/sid/>) and the PLANTS database. An  
 162 additional 15 species had their seed masses quantified using  
 163 seeds stored with herbaria sheets at the Michigan State  
 164 University Herbarium by N.G.S. The maximum height,  
 165 leaf size and seed mass data were all log transformed for  
 166 the downstream analyses given their highly skewed global  
 167 distributions.

### 168 Phylogenetic generalized least squares regression

169 We used pGLS regression to model the trait data for species  
 170 on one continent given their phylogenetic position and the  
 171 phylogenetic distribution of traits for species on the second  
 172 continent. A pGLS regression can incorporate the phyloge-  
 173 netic non-independence of data points by assuming a phylo-  
 174 genetic error structure given a model of trait evolution. In  
 175 the simplest case, a Brownian motion model of trait evolu-  
 176 tion can be assumed in which the error structure takes the  
 177 form of an untransformed phylogenetic variance–covariance  
 178 (VCV) matrix where the diagonal is the root to tip distance  
 179 and the off-diagonal elements are the amounts of shared  
 180 branch length between two taxa. This basic model can  
 181 become more flexible by fitting a model of trait evolution  
 182 given the data by transforming the phylogenetic VCV matrix  
 183 and finding the transformation that best fits the data (Swen-  
 184 son, 2014a,b). For example, if the data have no evident phy-  
 185 logenetic signal (i.e. non-independence) the transformation  
 186 of the off-diagonal values in the VCV matrix that would best  
 187 fit the data would be to multiply the values by zero. Simi-  
 188 larly, if the data are best explained by a Brownian motion

model the transformation that would best fit the data would  
 be to multiply the off-diagonal elements by one. The values  
 by which the off-diagonal elements are multiplied are  
 referred to as  $\lambda$ . We utilized maximum likelihood to estimate  
 the  $\lambda$  values (Pagel, 1999; Freckleton *et al.*, 2002) using the R  
 package ‘caper’ (<http://caper.r-forge.r-project.org/>) for each  
 trait on each continent and generated a GLS regression  
 model for that trait using the estimated phylogenetic error  
 structure (i.e. the transformed phylogenetic VCV matrix;  
 Swenson, 2014a,b). This model and the transformed VCV  
 matrix containing all species on both continents were then  
 used to predict the trait values of species on the other conti-  
 nent given the model from first continent. To assess the  
 degree to which the predicted species-level values were  
 related to the known values we regressed the predicted values  
 against the known values.

Next, the predicted values were then used to quantify the  
 mean and variance of traits in map grid cells on each conti-  
 nent as well as the multivariate functional dispersion (FDis)  
 and functional richness (FRic) in those grid cells. The FDis is  
 the mean distance of each species to the centroid of the mul-  
 tivariate trait space and the FRic is the volume of the multi-  
 variate trait space that an assemblage occupies (Laliberte &  
 Legendre, 2010). These values were then compared with the  
 known values using a regression.

A simple alternative to estimating the most likely  $\lambda$  values  
 for a given trait dataset and phylogeny is just to assume that  
 traits evolve under a Brownian motion model. For example,  
 a Brownian motion model could be assumed where the phylo-  
 genetic VCV matrix is left untransformed (i.e.  $\lambda = 1$ ). We  
 generated these models for each trait on each continent and  
 used the models and an untransformed phylogenetic VCV  
 matrix containing all species to predict the trait values on  
 the other continent. As with the previous analysis, we then  
 regressed predicted trait values for species against their  
 known values. Then, the predicted values were used to quan-  
 tify the mean and variance of traits in map grid cells on each  
 continent as well as the multivariate FDis and FRic in those  
 grid cells.

### Phylogenetic eigenvector regression

In addition to the two pGLS approaches used to predict trait  
 values, we utilized phylogenetic eigenvectors to predict trait  
 values, which assume no model of trait evolution – Brownian  
 motion or otherwise. To accomplish this, a phylogenetic dis-  
 tance matrix was computed from the phylogeny and used in  
 a principal coordinate analysis to generate phylogenetic  
 eigenvectors (Diniz-Filho *et al.*, 1998; Ramirez *et al.*, 2008;  
 Diniz-Filho *et al.*, 2011). The number of phylogenetic eigen-  
 vectors produced is equal to the number of species minus  
 one. A subset of eigenvectors must be selected for phyloge-  
 netic eigenvector regression because the use of all eigenvec-  
 tors leads to model saturation (Rohlf, 2001). We utilized an  
 iterative search for the subset of eigenvectors that reduces the  
 largest amount of autocorrelation in the residuals (Griffith &

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243 Peres-Neto, 2006; Diniz-Filho *et al.*, 2012). Specifically, as  
 244 new eigenvectors were added to the model for a single trait  
 245 on a single continent, residual autocorrelation was recalcu-  
 246 lated and the iterative search stopped until the residual auto-  
 247 correlation calculated using Moran's *I* was less than 0.05. The  
 248 selected eigenvector values for species on one continent were  
 249 then used as independent variables in a multiple linear model  
 250 with the data for a single trait from the same continent as  
 251 the dependent variable. This model was then projected onto  
 252 the values for the species on the other continent from the  
 253 same subset of eigenvectors. This process was repeated for  
 254 each trait to produce predicted trait values on one continent  
 255 given the trait data on the other continent and their phyloge-  
 256 netic eigenvector positions. The R package 'PVR' was used  
 257 for all phylogenetic eigenvector analyses (<http://cran.r-project.org/web/packages/PVR/>). Again, the predicted species-level  
 259 trait values were regressed onto the known values through  
 260 the origin and the coefficient of determination was recorded.  
 261 Next, the predicted trait values derived from this phyloge-  
 262 netic eigenvector approach were then used to quantify the  
 263 mean and variance of traits in map grid cells on each conti-  
 264 nent as well as the multivariate FDis and FRic in those grid  
 265 cells. These values were then compared with the known  
 266 values.

267 **Prediction error and climate**

268 Deviations of the predicted map grid cell values from the  
 269 'known' values may be linked to climate. We therefore per-  
 270 formed a series of ad hoc tests in which we first quantified  
 271 the deviation of the predicted values from the known values  
 272 (i.e. known value minus the predicted value) and correlated  
 273 these values with four climatic variables for the same grid  
 274 cell. Specifically, we used Pearson correlations to evaluate the  
 275 relationships between the deviations and mean annual tem-  
 276 perature, temperature seasonality, annual precipitation and

precipitation seasonality using climate maps from the World- 277  
 Clim database (Hijmans *et al.*, 2005) at a resolution of 2.5°. 278

279 **RESULTS**

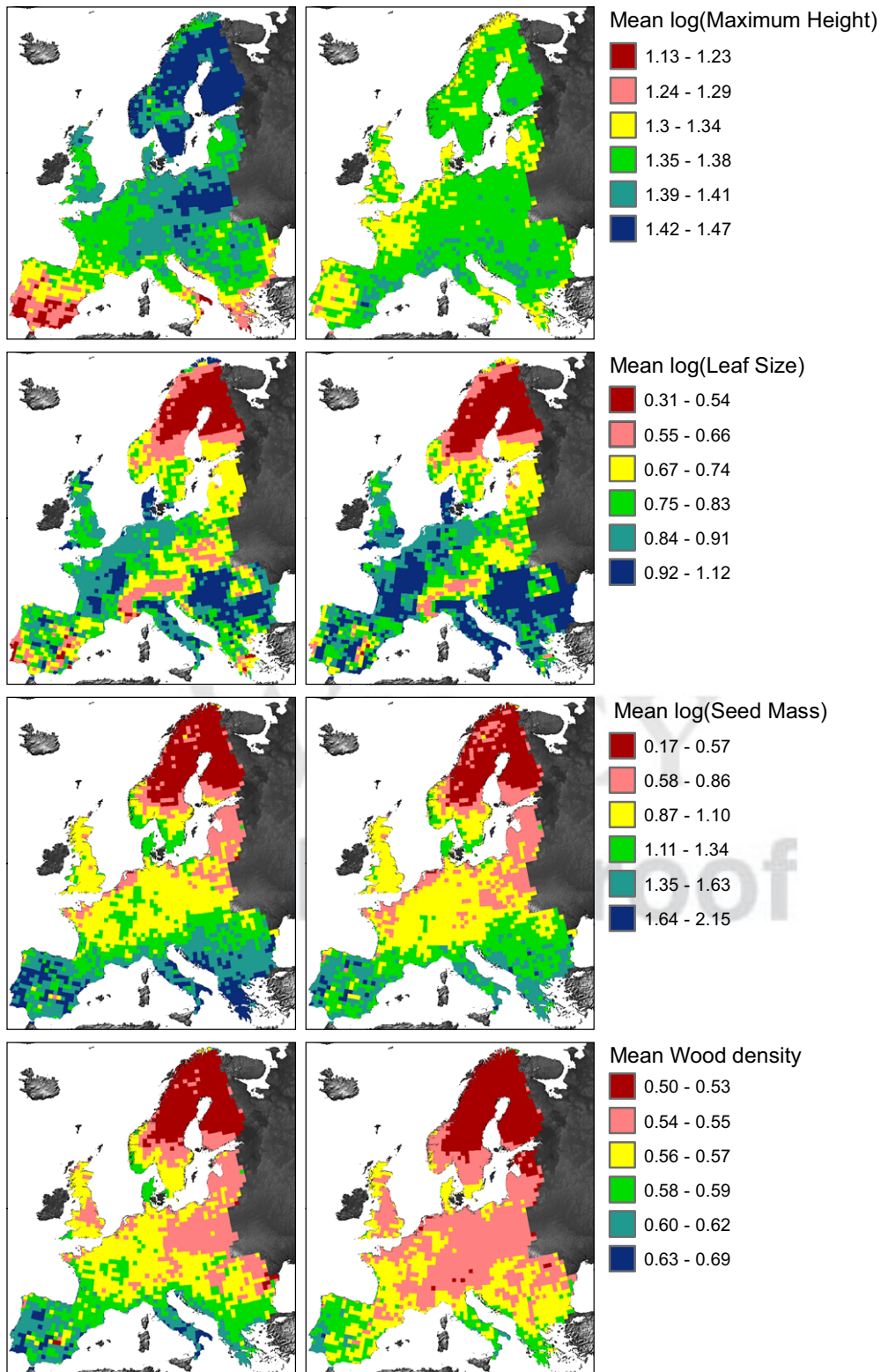
280 We utilized three phylogenetic imputation methods to pre- 280  
 dict the trait values of species in one region (eastern North 281  
 America or Europe) based upon their phylogenetic position 282  
 and the traits and the phylogenetic position of species in the 283  
 other region. We used the predicted values to map the mean 284  
 and variance of each trait and to estimate two multivariate 285  
 functional diversity indices in the map grid cells in each 286  
 region. We began by testing the pGLS regression with a fit 287  
 model of trait evolution. The predicted trait means, FDis and 288  
 FRic in map grid cells in the projection region based on trait 289  
 information in the calibration region and phylogenetic infor- 290  
 mation were typically highly correlated ( $r^2 > 0.60$ ; Table 1, 291 T1  
 Figs 1 & 2). The predicted trait variances in map grid cells 292 F1 F2  
 were also highly correlated with the known variances 293  
 ( $r^2 > 0.60$ ; Table 1). However, predictions of the mean and 294  
 variance of maximum height values for the map grid cells 295  
 were far weaker ( $r^2 \sim 0.1-0.3$ ) indicating that the lability in 296  
 the evolution of this trait prevented strong predictions even 297  
 when  $\lambda$  was estimated and used to fit the model. The geo- 298  
 graphical locations that were the most difficult to predict in 299  
 Europe were typically in the south-east (Figs 1 & 2). Simi- 300  
 larly, the more species-rich south-eastern portion of eastern 301  
 North America was the region hardest to predict, probably 302  
 due to the higher number of congeners and the greater num- 303  
 ber of species that may be distantly related from the dataset 304  
 used to build the statistical model. 305

306 To explore whether alternative phylogenetic prediction 306  
 frameworks provided similarly strong predictions we took 307  
 two additional approaches. First, we did not use maximum 308  
 likelihood to estimate  $\lambda$  values in the pGLS model. Rather, 309  
 we used the observed phylogenetic VCV matrix in the pGLS 310

**Table 1** We used phylogenetic generalized least squares (pGLS) regression to estimate a model of trait evolution ( $\lambda$ ) using the trait data from one continent to predict the trait values for species on the other continent. The table shows the intercept and slope of each regression with their standard errors (SE) and  $r^2$ . We also report the  $\lambda$  values estimated by our pGLS models where values closer to one indicate more phylogenetic signal and values closer to zero indicate less phylogenetic signal.

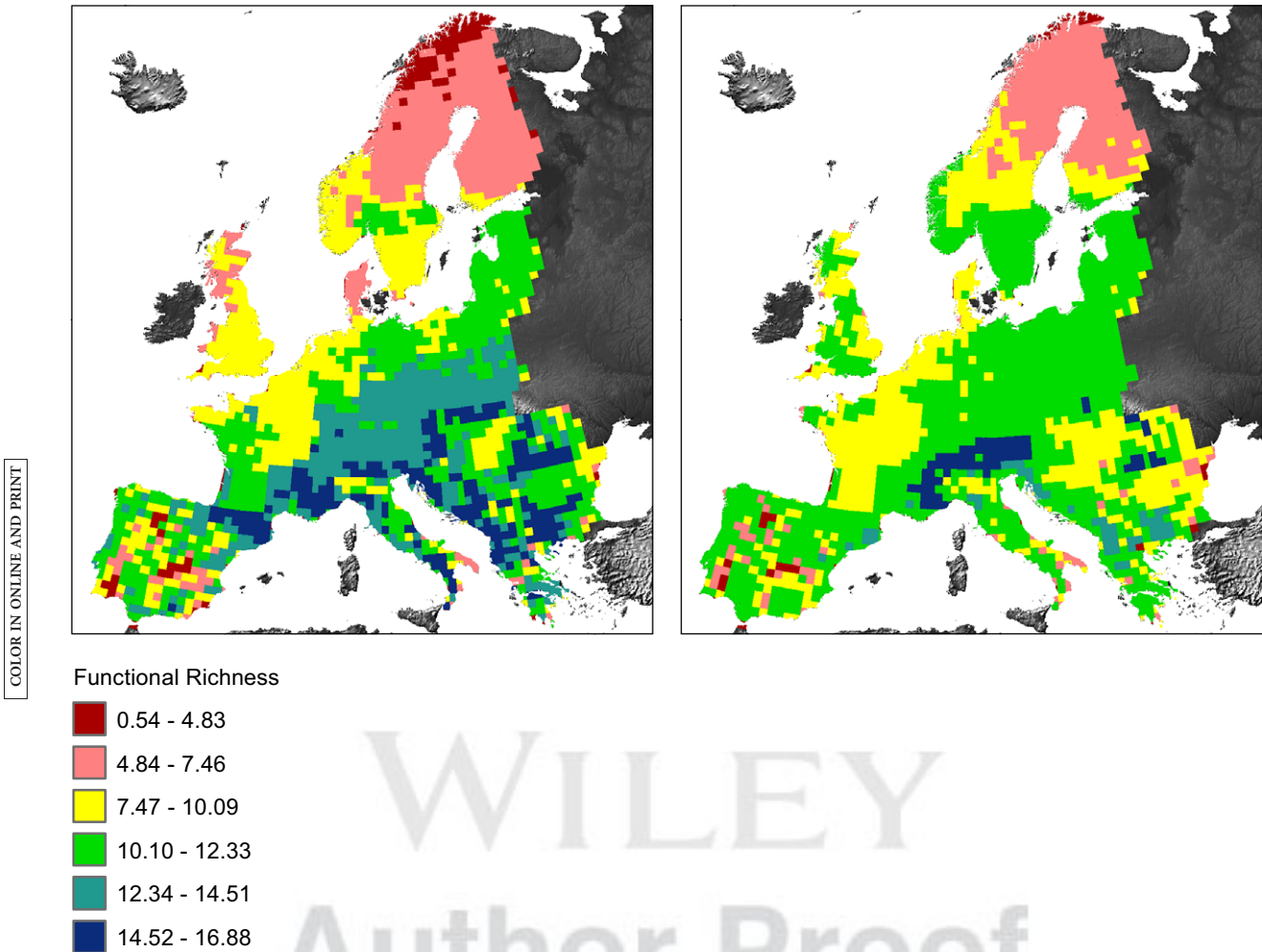
Map grid cell value	Eastern North America prediction of European traits						European prediction of eastern North American traits					
	Intercept	SE	Slope	SE	$r^2$	$\lambda$	Intercept	SE	Slope	SE	$r^2$	$\lambda$
Mean maximum height (m)	1.13	0.01	0.16	0.01	0.28	0.68	1.39	0.01	-0.02	0.01	0.10	0.65
Variance maximum height (m)	0.02	< 0.00	-0.07	0.01	0.14	0.68	0.02	< 0.00	-0.11	0.01	0.15	0.65
Mean leaf size (cm <sup>2</sup> )	-0.09	0.01	1.17	0.01	0.87	0.96	0.17	0.01	0.59	0.02	0.76	0.98
Variance leaf size (cm <sup>2</sup> )	-0.03	< 0.00	0.94	0.01	0.92	0.96	-0.02	0.01	0.63	0.01	0.88	0.98
Mean seed mass (g)	0.13	< 0.00	0.79	< 0.00	0.97	0.99	0.41	0.01	0.82	0.01	0.92	0.89
Variance Seed Mass (g)	0.16	0.01	0.64	0.01	0.87	0.99	0.59	0.01	0.30	0.02	0.42	0.89
Mean wood density (g cm <sup>-3</sup> )	0.01	< 0.00	0.97	0.01	0.93	0.85	-0.02	0.01	1.04	0.02	0.99	0.85
Variance wood density (g cm <sup>-3</sup> )	0.01	< 0.00	0.12	0.01	0.30	0.85	< 0.00	< 0.00	0.20	0.02	0.38	0.85
Functional dispersion	0.46	0.03	0.77	0.01	0.44	-	0.10	0.03	1.10	0.01	0.84	-
Functional richness	3.67	0.10	0.62	0.01	0.66	-	4.29	0.16	0.81	0.02	0.80	-

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**Figure 1** The known (left) and predicted (middle) trait means in map grid cells for European trees. Deviations (right) where the predicted values were subtracted from the known value are also plotted. The top row is mean maximum height (log m), the second row is mean leaf size (log cm<sup>2</sup>), the third row is mean seed mass (log g) and the fourth row is mean wood density (g cm<sup>-3</sup>). The predicted values were generated by fitting a model of trait evolution for maximum height, leaf size, seed mass and wood density for eastern North American trees and using that model to predict the trait values of European tree species based on their phylogenetic position. The colour legends are provided on the right side of each row with the top legend corresponding to the maps in the first two columns (i.e. the trait means) and the bottom legend to the map in the last column (i.e. the deviations).

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**Figure 2** The multivariate functional richness (FRic) quantified using the known trait data (left) and the predicted trait data (right) for European trees. The predicted values were generated by fitting a model of trait evolution for maximum height, leaf size, seed mass and wood density for eastern North American trees and using that model to predict the trait values of European tree species based on their phylogenetic position. The known and predicted FRic values are highly correlated ( $r^2 = 0.964$ ) with a lower than expected root mean squared error (RMSE = 1.944;  $P < 0.05$ ).

311 model, effectively assuming a  $\lambda$  value of one (i.e. Brownian  
 T2 312 motion trait evolution) for every trait dataset (Table 2). Sec-  
 313 ond, we utilized a phylogenetic eigenvector regression that  
 T3 314 does not fit a model of trait evolution (Table 3). The results  
 315 from both pGLS approaches were qualitatively similar (Tables  
 316 1 & 2) where strong predictions were possible for most traits,  
 317 with the notable exception of maximum height. The phyloge-  
 318 netic eigenvector predictions were less robust, with some  
 319 traits having strong predictions (e.g. leaf area and seed  
 320 mass); wood density and maximum height predictions were  
 321 less strong (Table 3).  
 322 Lastly, we quantified the correlation between four climatic  
 323 variables and deviations of the predicted values from known  
 324 values for map grid cells on both continents. We found that  
 325 deviations were nearly always correlated with the four  
 326 climatic variables (Tables S1 & S2 in the Supporting Infor-  
 327 mation). The correlations were generally stronger for  
 328 temperature-related variables than for precipitation-related

variables. The geographical signature in the deviations for 329  
 Europe can be seen in Fig. 1, indicating that in the study sys- 330  
 tem the major deviations generally occur at the extremes of 331  
 latitude. 332

## DISCUSSION 333

Mapping the distribution and diversity of plant function on 334  
 continental scales is a fundamental goal in biogeography and 335  
 ecosystem ecology (Reich, 2005; Swenson & Weiser, 2010; 336  
 Swenson *et al.*, 2012). A key limitation to progress is that 337  
 most large plant trait databases are highly sparse (Kattge 338  
*et al.*, 2011) so probably making most efforts at functional 339  
 trait mapping prone to large error. While waiting for more 340  
 data to accumulate, a pragmatic way forward may be to 341  
 impute or estimate the missing trait values in existing data- 342  
 bases. These estimates could be strengthened by incorporat- 343  
 ing phylogenetic information (Swenson, 2014a; Schrodte *et al.*, 344

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**Table 2** In this table we do not estimate a model of trait evolution, rather we assume a Brownian motion model of trait evolution ( $\lambda = 1$ ) and phylogenetic generalized least squares. The predicted trait values and species distribution maps were then used to calculate the predicted mean and variance of each trait value and the predicted multivariate functional dispersion and functional richness value in map grid cells on each continent. The predicted mean, variance, functional dispersion and functional richness values were regressed onto the known values. The table gives the intercept and slope of each regression with their standard errors (SE) and the  $r^2$ .

Map grid cell value	Eastern North America prediction of European traits					European prediction of eastern North American traits				
	Intercept	SE	Slope	SE	$r^2$	Intercept	SE	Slope	SE	$r^2$
Mean maximum height (m)	1.13	0.01	0.162	0.01	0.18	1.33	0.01	0.026	0.01	0.11
Variance maximum height (m)	0.02	< 0.00	-0.068	0.01	0.14	0.02	< 0.00	-0.118	0.01	0.18
Mean leaf size (cm <sup>2</sup> )	-0.09	0.01	1.174	0.01	0.88	0.17	0.01	0.604	0.01	0.77
Variance leaf size (cm <sup>2</sup> )	-0.03	< 0.00	0.942	0.01	0.93	-0.02	0.01	0.631	0.01	0.88
Mean seed mass (g)	0.13	< 0.00	0.794	< 0.00	0.98	0.28	0.01	0.907	0.01	0.91
Variance seed mass (g)	0.10	0.01	0.641	0.01	0.88	0.68	0.02	0.316	0.02	0.41
Mean wood density (g cm <sup>-3</sup> )	0.01	< 0.00	0.973	0.01	0.93	-0.03	0.01	1.076	0.02	0.99
Variance wood density (g cm <sup>-3</sup> )	0.01	< 0.00	0.117	0.01	0.26	0.01	< 0.00	0.243	0.02	0.30
Functional dispersion	0.46	0.02	0.767	0.01	0.44	0.07	0.02	1.121	0.01	0.88
Functional richness	3.67	0.09	0.619	0.01	0.67	4.16	0.15	0.786	0.01	0.81

2015). This is because some plant functional traits of interest are known to have a phylogenetic signal in global datasets (e.g. Moles *et al.*, 2005; Swenson & Enquist, 2007). The goal of the present work was to implement and test the ability of phylogenetic imputation methods to predict the distribution and diversity of plant functional traits on continental scales.

Here, we have shown that robust predictions of individual trait distributions and the overall functional diversity within map grid cells can be predicted among novel continental settings simply by taking advantage of the phylogenetic signal in trait data from another continent. The three approaches to phylogenetic imputation used here all were able to predict a large amount of the variance in trait distributions at the species and map grid cell levels (Tables 1–3). However, the

two pGLS regression-based approaches explained more variance than phylogenetic eigenvector regression-based methods. Thus, even when setting aside conceptual debates regarding eigenvector approaches (e.g. Rohlf, 2001), these methods tended to perform well, but not as well as pGLS. The two pGLS approaches yielded similar results in this study, but it is expected that in many cases trait evolution will not as closely approximate a Brownian motion model and the pGLS approach fitting a  $\lambda$  value will be more reliable. More work is needed, using larger empirical and simulated datasets, (Swenson, 2014a) to confirm or reject this general recommendation.

The phylogenetic imputation methods were able to make strong predictions of the spatial distribution of traits, but the

**Table 3** We used phylogenetic eigenvector regression using the trait data from one continent to predict the trait values for species on the other continent. The predicted trait values and species distribution maps were then used to calculate the predicted mean and variance of each trait value and the predicted multivariate functional dispersion and functional richness value in map grid cells on each continent. The predicted mean, variance, functional dispersion and functional richness values were regressed onto the known values through the origin. This table shows the intercept and slope of each regression with their standard errors (SE) and the  $r^2$ .

Map grid cell value	Eastern North America prediction of European traits					European prediction of eastern North American traits				
	Intercept	SE	Slope	SE	$r^2$	Intercept	SE	Slope	SE	$r^2$
Mean maximum height (m)	1.26	0.01	0.06	0.01	0.11	1.33	0.01	0.01	0.01	0.08
Variance maximum height (m)	0.01	< 0.00	-0.05	0.01	0.11	0.01	< 0.00	-0.06	0.01	0.11
Mean leaf size (cm <sup>2</sup> )	-0.17	0.01	1.16	0.01	0.75	0.27	0.01	0.53	0.02	0.63
Variance leaf size (cm <sup>2</sup> )	0.02	0.01	0.76	0.01	0.46	0.41	0.02	0.43	0.02	0.46
Mean seed mass (g)	0.36	< 0.00	0.73	< 0.00	0.96	1.03	0.01	0.43	0.02	0.52
Variance seed mass (g)	-0.08	0.01	0.60	0.01	0.81	0.65	0.03	0.18	0.01	0.11
Mean wood density (g cm <sup>-3</sup> )	0.22	0.01	0.60	0.02	0.51	0.03	0.02	0.96	0.04	0.93
Variance wood density (g cm <sup>-3</sup> )	< 0.00	< 0.00	0.04	0.01	0.15	0.01	< 0.00	-0.06	0.01	0.09
Functional dispersion	0.09	0.03	0.85	0.01	0.44	0.21	0.03	0.92	0.02	0.69
Functional richness	0.44	0.15	0.90	0.01	0.65	-1.26	0.56	1.88	0.02	0.64

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373 northern- and southernmost portions of both regions were  
374 where the methods performed worst (Figs 1 & 2). This is  
375 particularly evident when we consider the strong relation-  
376 ships between temperature variables and the deviation of pre-  
377 dicted values from known values (Tables S1 & S2). One  
378 reason for this may be the tendency of the methods to  
379 under-represent trait divergences due to habitat differences  
380 within a clade and an over-averaging of trait data leading to  
381 higher deviations in more extreme climates within clades.  
382 Future work may be able to remedy this bias by either incor-  
383 porating climatic information into the species-level trait pre-  
384 dictions or adjusting predicted species-level values in map  
385 grid cells or the assemblage-level trait or diversity values  
386 based upon climate, but such work is beyond the scope of  
387 the present paper. A second reason is that these regions con-  
388 tain a greater number of species from different parts of the  
389 phylogenetic tree. In other words, the distance between a  
390 data point used to build the statistical model and a species  
391 in these regions will increase. This is particularly the case  
392 when building a model on one continent and projecting it to  
393 another where it is likely that many genera in the species-  
394 rich regions on the continent to be predicted are not found  
395 on the continent used to build the model.

396 This study focused on four functional traits commonly  
397 used in trait-based ecology and readily available in the litera-  
398 ture. Two of these traits, wood density and seed mass, are  
399 known to have a great deal of phylogenetic signal (Moles  
400 *et al.*, 2005; Swenson & Enquist, 2007), meaning that phylo-  
401 genetic imputation methods are likely to be very successful.  
402 Indeed, we found this to be the case at the species and map  
403 grid cell levels (Tables 1–3). The phylogenetic signal in the  
404 other two traits, maximum height and leaf size, has not been  
405 as well scrutinized in the literature at global scales. Maximum  
406 height was found to have much less phylogenetic signal than  
407 the other traits, but leaf size had a similar degree of phyloge-  
408 netic signal to seed mass and wood density (Table 1). The  
409 outcome of this was that predictions of maximum height dis-  
410 tributions were far less reliable than those of leaf size distri-  
411 butions (Tables 1–3).

#### 412 **Considerations for future implementation of** 413 **phylogenetic imputation**

414 It may seem surprising that our phylogenetically based  
415 approach is able to predict the observed geographical pat-  
416 terns so strongly. We expect that some of this success is due  
417 to the fact that the two tree floras are very similar in their  
418 familial and generic compositions. Thus, the average phyloge-  
419 netic distance between a training trait data point and a pre-  
420 dicted trait data point is relatively low and represents  
421 perhaps a best-case scenario. In other words, projecting the  
422 traits of another flora with a very different phylogenetic com-  
423 position (e.g. the Amazon) from European data would be  
424 likely to result in much more error. Indeed, we found evi-  
425 dence of this to a smaller degree when we consider that less  
426 variation in eastern North America could be predicted using

the smaller European flora than vice versa (e.g. Table 1). 427 AQ6  
428 Additionally, the methods used are regressions and extrapola-  
429 tions of these models, so will more likely than not introduce  
430 large errors. In the present study, the bounds of the data in  
431 each region are roughly similar, but if one region lacked, for  
432 example, gymnosperms there would be a highly increased  
433 potential for error. Taken together, future work will have to  
434 closely consider the phylogenetic compositions of the training  
435 data set and the species set to be predicted. Some of the  
436 potential for error could be mitigated by using the largest  
437 trait datasets available (e.g. Kattge *et al.*, 2011; Schrodte *et al.*,  
438 2015) such that phylogenetic extrapolation does not occur  
439 and the predicted trait values can stay within reasonable  
440 bounds.

441 Another consideration arising from this study is that we  
442 only considered four traits that, while being of interest to  
443 ecologists, do not represent the entirety of the traits that  
444 ecologists are interested in mapping. For example, earth sys-  
445 tem modellers are likely to be more interested in leaf gas  
446 exchange rates that may be highly variable within families  
447 and genera (i.e. have little phylogenetic signal; see van Bode-  
448 gom *et al.*, 2012). Such traits may approximate the situation  
449 we encountered with maximum height where predictions are  
450 not as strong, and this would propagate error once aggre-  
451 gated into things like global dynamic vegetation models.  
452 Thus, an important question will be the degree to which the  
453 error introduced via phylogenetic imputation is less or more  
454 than the error introduced by lumping species into a few dis-  
455 crete functional types.

456 Next, the present study found strong relationships between  
457 climate and deviations from predictions. Each of the meth- AQ7  
458 ods used could incorporate climatic information by quantifying  
459 the average climate for each species and using this  
460 information as an additional independent variable in the  
461 model, such that phylogenetic signal and trait–climate rela-  
462 tionships are simultaneously used to predict missing trait val-  
463 ues. It is expected that such models will strengthen trait  
464 predictions, particularly when phylogenies with no resolution  
465 within genera are utilized. An alternative approach could be  
466 adjusting post hoc the grid cell values for assemblages by cli-  
467 mate, but this approach may be more arbitrary and unreli-  
468 able. More detailed future models may also seek to model  
469 population-level response to climate hierarchically, which  
470 may help refine predictions of traits that are very sensitive to  
471 local abiotic conditions (e.g. gas exchange). However, to our  
472 knowledge, such phylogenetically explicit methods that model  
473 trait evolution along branch lengths have not yet been  
474 developed.

475 Lastly, it is worth highlighting again that the proposed  
476 methods are meant to serve as a pragmatic approach to esti-  
477 mating trait values given the current circumstances. Without  
478 a doubt we would prefer that trait values were actually meas-  
479 ured than predicted, and future trait collection campaigns,  
480 particularly in under-sampled regions like the tropics, should  
481 remain a priority. Further, as previously noted (see Swenson,  
482 2014a), while the biases or errors introduced by phylogenetic



483 imputation may be tolerable on very large scales, using  
 484 imputed values for local-scale studies or community ecology  
 485 would be likely to introduce levels of error that would not be  
 486 tolerable. Thus, we are not recommending the use of these  
 487 methods for trait-based community ecology.

## 488 CONCLUSIONS

489 In recent years plant ecologists and evolutionary biologists  
 490 have made tremendous advances by generating and analysing  
 491 large plant trait databases (Kattge *et al.*, 2011) and large phy-  
 492 logenetic trees (Webb & Donoghue, 2005). We suggest that  
 493 these advances can now be leveraged to produce phylogeneti-  
 494 cally based predictions of the continental-scale distribution  
 495 and the diversity of plant function, even into areas with  
 496 novel sets of species. This predictive power will be crucial in  
 497 a future where climate change and species introductions will  
 498 increasingly generate novel assemblages. Importantly these  
 499 predictions may be the most pragmatic way for ecosystem  
 500 modellers to incorporate functional diversity within and  
 501 among map grid cells into their models and move beyond  
 502 using a singular plant functional type to represent all vegeta-  
 503 tion within a region, and to do so even for less-studied  
 504 regions with many species for which we have little direct trait  
 505 information.

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**SUPPORTING INFORMATION**

Additional supporting information may be found in the  
 online version of this article at the publisher’s web-site:

**Table S1** Pearson correlation coefficients for eastern North  
 America between the deviation of the predicted values from  
 the known value where the deviation is calculated as the  
 predicted subtracted from the observed.

**Table S2** Pearson correlation coefficients for Europe between  
 the deviation of the predicted values from the known value  
 where the deviation is calculated as the predicted subtracted  
 from the observed.

**BIOSKETCH**

**Nathan G. Swenson** is a plant biologist interested in  
 the distribution of biodiversity. His work focuses on  
 patterns of species and functional and phylogenetic  
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 that underlie them.

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